

Hardware and Software Installation

- Screw in the Bluetooth antenna; or connect the instrument using the USB cable
- Connect the power supply to the instrument and turn the instrument on
A blue light on the front of the instrument illuminates to indicate it is powered on
- Install the micPCR software from the provided USB key
The USB key also contains an electronic copy of the user manual
- Open the micPCR software
The instrument will be automatically detected and appear in the top right corner
- For the latest software updates visit and register at www.biomolecularsystems.com/customer-resources/

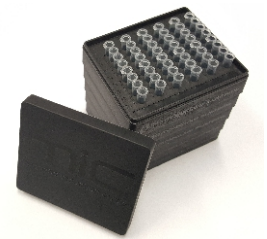


Reaction Setup

- Load reactions into the pre-racked **Mic Tubes** and use the **Capping Tool** to help with fitting caps properly
- Insert the tubes into the rotor, ensuring the tab is aligned with the marks on the rotor



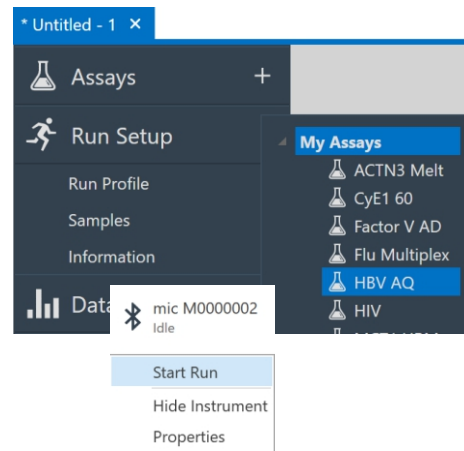
To achieve optimum temperature uniformity, it is very important to insert tubes loaded with water (at reaction volume) into all of the unused positions on the rotor. See reverse for more details.



- Secure the tubes into the rotor with the **Tube Clamp**

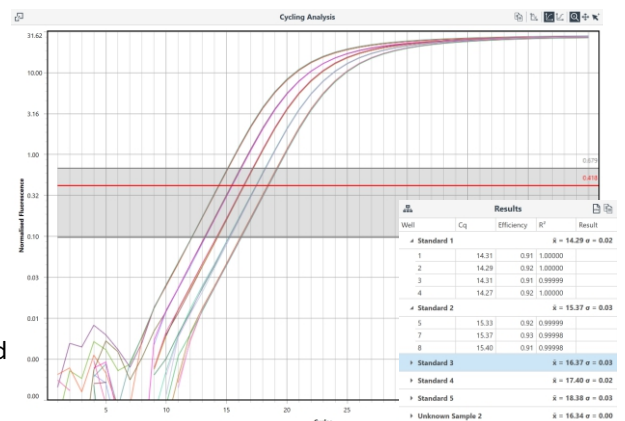
Starting a Run

- Select **New Run** from the toolbar
- Add (**Assays +**) an assay to the run from the assay list
Additional assay's are easily created by selecting **New Assay** from the toolbar
- Click on the instrument (top right of toolbar) and select **Start Run**
- Confirm the reaction volume is correct and select **Start**
A green light on the front of the instrument will indicate the run has begun
- Before analysis, fill in the sample information on the **Samples** page
- Drag and drop the assay for each well onto the sample editor
Assay's are used to link samples together for later analysis



Analysis

- Select the required analysis to the run from the options provided
 - Cycling
 - Melt
 - High Resolution Melt (optional)
 - Absolute Quantification (includes Standard Curve)
 - Allelic Discrimination
 - Relative Quantification
- Select the target to analyse and configure the parameters as required
- To learn more about our software visit www.mic-qpcr/tutorials

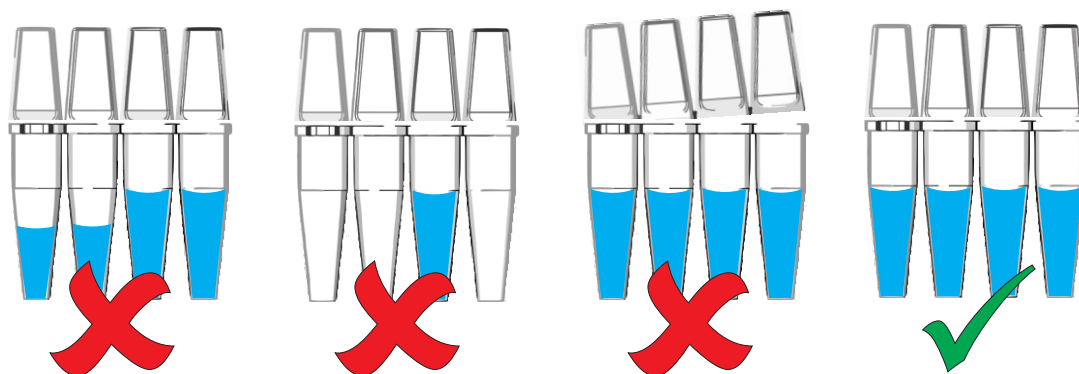


Water Tubes

No tubes, empty tubes and tubes with different volumes of liquid all have different thermal loads on the metal rotor. Variations in thermal load around the rotor can cause significant thermal gradients both at static temperatures and during ramping, resulting in increased variability in results.

For the best thermal uniformity between samples always follow these simple rules:

- Ensure tubes are loaded into every position around the rotor
- Load unused tubes with water, at the reaction volume
- Check all tubes are firmly pushed into the rotor and the caps are securely fitted
- Use the tube clamp to keep tubes in the rotor during the run



It is very important that all tubes within the rotor contain the same volume of liquid during a run. Empty tubes should be loaded with water.

As the oil overlay prevents evaporation, these *Water Tubes* can be stored and reused for over a week.

